

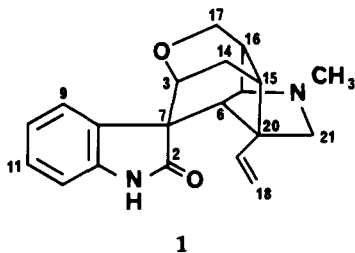
STUDIES ON THE NMR SPECTROSCOPIC PROPERTIES OF GELSEMINE—REVISIONS AND REFINEMENTS

YEH SCHUN and GEOFFREY A. CORDELL*

Program for Collaborative Research in the Pharmaceutical Sciences, College of Pharmacy, University of Illinois at Chicago, Chicago, Illinois 60612

Gelsemium sempervirens (L.) Jaume St.-Hilaire has been used in medicine for its antispasmodic and analgesic properties. It was at one time recognized in several pharmacopoeias and found use in the treatment of influenza (1,2). For toxicity reasons, it is no longer included in the current pharmacopoeias.

The principal alkaloid, gelsemine, was first isolated by Wormley in 1870, and its physiological properties in vivo have been studied for many years (3-8). It is not the most potent alkaloid in this series thus far known; gelsemicine has that distinction. The structure of gelsemine (**1**) was established through



X-ray crystallographic analysis (9) more than 90 years after its initial discovery. Mass spectrometry proved to be a useful tool for the identification of gelsemine and its analogs (10) and 220 MHz ^1H -nmr (11) and ^{13}C -nmr spectra (11, 12) of gelsemine (**1**) have been described.

As part of our program on the isolation of plant anticancer agents, we found that extracts of *G. sempervirens* displayed cytotoxic activity in the KB or P-388 test system (13). In order to assist in future studies of alkaloids in this series, we decided to reestablish the ^1H - and ^{13}C -nmr assignments obtained previously.¹

¹Gelsemine was actually the first alkaloid whose ^{13}C -nmr spectrum was assigned (12).

Although most of the assignments in the ^1H - and ^{13}C -nmr spectra (Table 1) compare favorably with those derived earlier (11,12), a number of revisions were found to be necessary.

Figure 1 shows the homonuclear 2-D correlation spectrum of gelsemine in the range 1.9-4.1 ppm. It is clear that the original assignments for H-14a, H-14e and H-15 and H-16 should be revised. Thus, the signal at 2.43 ppm can now be attributed to H-16, rather than H-14a, since it is the only proton with coupling to the H-17 methylene protons. Consequently, the signal at 37.93 ppm could now be ascribed to C-16. The axial proton at C-14, coupling to both H-3 ($J=2.8$ Hz) and H-14e ($J=14.2$ Hz) but not to H-15, was assigned to the signal at 2.83 ppm. This is also clearly shown in the heteronuclear correlation spectrum of gelsemine, in which this signal correlates with that for C-14, which is the most upfield carbon at 22.67 ppm. The equatorial proton at C-14 appears as a doublet of doublets of doublets ($J=14.2, 6.1, 3.2$ Hz), as expected from Dreiding models where only the equatorial proton is predicted to couple with H-15. Revision in the assignment of H-15 is also necessary, though its precise chemical shift and multiplicity remain obscured below one of the C-21 methylene protons. The chemical shift is perhaps more clearly seen in Figure 1 where the coupling to H-14e is apparent. The corresponding carbon chemical shift is 35.54 ppm.

The only remaining protons to be discussed are therefore those on C-21. Assignment of these protons was made possible through the observation of long-range coupling between one of these

TABLE 1. Original and Revised Proton and ^{13}C -nmr Assignments of Gelsemine (1)^a

Proton	Original ^b Chemical Shift	Multiplicity	Revised ^d Chemical Shift	Multiplicity	Carbon	Original ^e	Revised ^f
H-3	3.79	d, 2.8	3.81	s	C-2	179.0	179.25
H-5	3.47	s	3.46	s	C-3	69.2	69.24
H-6	1.97	s	1.98	s	C-5	71.7	71.81
H-9	7.43	dd, 7.5, 2.0	7.40	d, 7.6	C-6	40.2	50.47
H-10	6.97	ddd, 7.5, 7.5, 2.0	6.99	ddd, 7.2, 7.2, 1.0	C-7	53.7	53.96
H-11	7.15	ddd, 7.5, 7.5, 2.0	7.17	ddd, 7.2, 7.2, 1.0	C-8	131.8	131.74
H-12	6.65	dd, 7.5, 2.0	6.74	d, 7.6	C-9	127.7 ^g	127.97
H-14a	2.37	d, 8.0	2.83	dd, 14.2, 2.8	C-10	121.4	121.41
H-14e	ca. 2.0	d, 8.0	2.01	ddd, 14.2, 6.1, 3.2	C-11	128.0 ^e	127.70
H-15	2.83	dd, 14.5, 2.8	ca. 2.30		C-12	108.7	108.80
H-16	ca. 2.30	ddd, 14.5, 2.0, 2.0	2.43	bd d, 8.3	C-13	140.3	140.52
H-17a	3.91	dd, 11.0, 2.0	3.92	dd, 11.0, 2.0	C-14	22.6	22.67
H-17e	4.10	dd, 11.0, 2.0	4.11	dd, 11.0, 2.0	C-15	37.8	35.54
H-18c	4.95	dd, 18.0, 1.8	4.95	dd, 17.8, 1.1	C-16	35.6	37.93
H-18t	5.09	dd, 11.0, 1.8	5.09	dd, 11.0, 1.1	C-17	61.1	61.33
H-19	6.28	dd, 18.0, 11.0	6.26	dd, 17.8, 11.0	C-18	111.9	111.89
H-21 exo	2.78 ^c	d, 10.0	2.78	d, 10.4	C-19	138.5	138.50
H-21 endo	2.32 ^c	d, 10.0	2.32	d, 10.4	C-20	53.7	53.82
N-CH ₃	2.24	s	2.25	s	C-21	65.9	65.88
NH	—		8.71	s	N-CH ₃	50.4	40.40

^a CDCl_3 was used as a solvent; chemical shifts in ppm; coupling constants in Hz.

^bObtained at 220 MHz, data from Wenkert *et al.* (12).

^cAssignments may be reversed.

^dObtained at 360 MHz.

^eObtained at 55.51 MHz, data from Wenkert *et al.* (12).

^fObtained at 90.84 MHz.

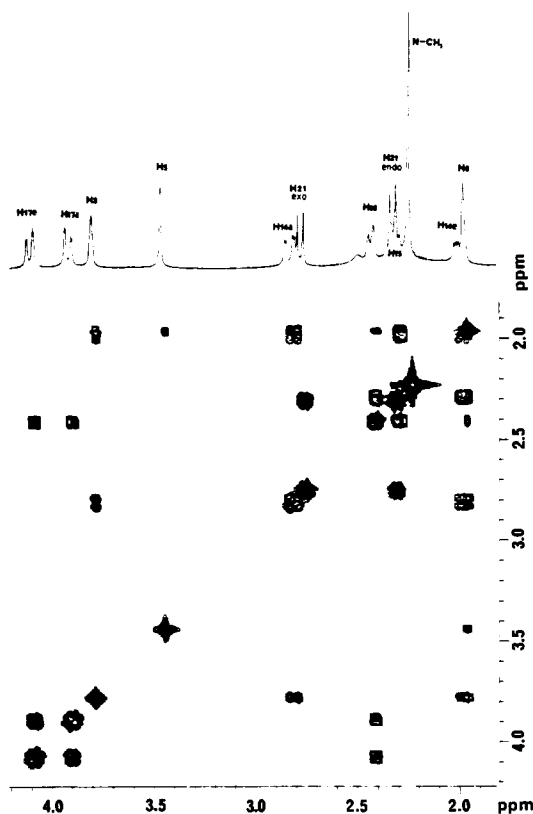


FIGURE 1. Two-dimensional correlation spectrum of gelsemine (1) in the range 1.9–4.1 ppm.

(2.78 ppm) and H-5 at 3.46 ppm. Molecular models indicate that this should be the *exo* (α) proton at C-21, leaving the doublet at 2.32 ppm to be assigned to the *endo* proton. Unfortunately, the close chemical shifts of H-15 and H-21 *endo* preclude observation of any nOe between these protons. However, a NOESY experiment did reveal an interaction between H-6 and H-21 *exo*, which substantiated the proton assignments. The N-CH₃ displayed nOe effects with both H-6 and H-16, as well as with both 21-protons, suggesting that inversion of the lone pair occurs with ease.

In the ¹³C-nmr spectrum, besides the revisions indicated previously, the most significant correction is for C-6 and the N-CH₃ group, whose assignments must now be reversed; C-6 at 50.47 ppm and N-CH₃ at 40.4 ppm. The heteronuclear COSY spectrum also allowed the assignment of C-9 (127.97 ppm) and C-11 (127.70 ppm) to be made unambiguously based on their proton multiplicities.

Two quaternary carbons, C-7 and C-20, also have very close chemical shifts. They were distinguished through a SINEPT experiment (14) irradiating H-9 and H-18. The signal at 53.96 ppm was selectively enhanced following irradiation of H-9, and the resonance at 53.82 ppm was enhanced on irradiating H-18. These signals are therefore C-7 and C-20, respectively.

EXPERIMENTAL

GENERAL EXPERIMENTAL PROCEDURES.—The ¹H-nmr spectra were recorded on a Nicolet Model NMC 360 (360 MHz) instrument with a Nicolet Fourier-Transform attachment using CDCl₃ as the solvent and TMS as an internal standard. The ¹³C-nmr spectrum was obtained in CDCl₃ at 90.84 MHz on the same instrument.

ISOLATION OF GELSEMINE (1).—Gelsemine (1) was isolated from the stem material of *G. sempervirens* through MeOH extraction, acid-base workup and column chromatography as described in a subsequent publication (15).

ACKNOWLEDGMENTS

This work was supported, in part, by Grant CA-20164 from the Division of Cancer Treatment, National Institutes of Health, Bethesda, MD. We gratefully acknowledge the Nuclear Magnetic Resonance Laboratory of the Research Resources Center, University of Illinois at Chicago, for providing the equipment and assistance necessary to conduct this study. The advice of Ms. L.-J. Lin in the establishment of the heteronuclear correlation spectroscopy experiment is also gratefully acknowledged.

LITERATURE CITED

1. T.Q. Chou, K.K. Chen, C. Pak, and H.C. Hou, *Chinese J. Physiol.*, **5**, 131 (1931).
2. O. Leeser, *J. Am. Inst. Homeopathy*, **29**, 730 (1936).
3. G. Tamba, *Acta Scholae. Med. Kyoto*, **4**, 85 (1921).
4. B.V. Christensen and L.G. Graming, *J. Am. Pharm. Assoc.*, **27**, 1208 (1938).
5. A. Risi, *Z. Biol.*, **99**, 446 (1939).
6. F.G. Henderson and K.K. Chen, *J. Am. Pharm. Assoc.*, **32**, 178 (1943).
7. L.S. Wu, *Bull. Natl. Formulary Comm.*, **15**, 68 (1947).
8. O. Eichler, F. Hertle, and I. Staib, *Arzneimittel-forsch.*, **7**, 349 (1957).
9. W.H. Orgell, *Lloydia*, **26**, 36 (1963).
10. F.M. Lovell, R. Pepinsky, and J.C. Wilson, *Tetrahedron Lett.*, (4), 1 (1959).
11. E. Wenkert, C.J. Chang, D.W. Cochran, and R. Pellicciari, *Experientia*, **28**, 377 (1972).
12. E. Wenkert, C.J. Chang, A.O. Clouse, and D.W. Cochran, *Chem. Commun.*, 961 (1970).
13. R.I. Geran, N.H. Greenberg, M.M. McDonald, A.M. Schumacher, and B.J. Abbott, *Cancer Chemother. Rep.*, **3**(2), 1 (1972).
14. A. Bax, *J. Mag. Reson.*, **57**, 314 (1984).
15. Y. Schun and G.A. Cordell, *J. Nat. Prod.*, (in preparation).

Received 12 April 1985